भारतीय मानक

IS 1909 : 2023

Indian Standard

भारतीय करी पाउडर — विशिष्टि

(दूसरा पुनरीक्षण)

Indian Curry Powder — Specification

(Second Revision)

ICS 67.220.10

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Price Group 7

Spices and Condiments Sectional Committee, FAD 09

FOREWORD

This Indian Standard (Second Revision) was adopted by the Bureau of Indian Standards, after the draft finalized by the Spices and Condiments Sectional Committee had been approved by the Food and Agriculture Divisional Council.

Indian curry powder is a blend of numerous spices with varying formulae to suit different tastes. Indian curry powders are generally used in cooking to impart taste and flavour to a variety of culinary preparations.

This Indian Standard was first issued in 1961 and revised in 1992 as a broad band of varieties of curry powders were being marketed in the country. With a view to enable monitoring of quality of Indian curry powders, it was considered imperative to revise the specification. Amendments were issued to the standard in 1994 and 2017 to keep the standard relevant to trade practices. This second revision is being carried out to align the standard with latest regulations laid under *Food Safety and Standards Act*, 2006 and the modifications incorporated in this revision are given below:

a) The microbiological requirements have been aligned with regulatory requirements;
b) Limits of aflatoxins have been incorporated;
c) Methods of test have been updated; and
d) References have been updated.

In the preparation of this standard due consideration has been given to the *Food Safety and Standards Act*, 2006 and the Rules and the Regulations framed thereunder and the *Legal Metrology (Packaged Commodities) Rules*, 2011. However, this standard is subject to restrictions imposed under these rules, wherever applicable.

The composition of the technical committee responsible for formulation of this standard listed in Annex E.

For the purpose of deciding whether a particular requirement of this standard is complied with, the final value, observed or calculated expressing the result of a test or analysis, shall be rounded off in accordance with IS 2 : 2022 ‘Rules for rounding off numerical values (*second revision*)’. The number of significant places retained in the rounded off value should be the same as that of the specified value in this standard.

Indian Standard

INDIAN CURRY POWDER — SPECIFICATION

(Second Revision)

1 SCOPE

This Indian Standard prescribes the requirements, method of sampling and tests for Indian curry powder.

2 REFERENCES

The following standards listed in Annex A contain provisions which, through reference in this text, constitute provisions of this standard. At the time of publication, the editions indicated were valid. All standards are subject to revision and parties to agreements based on this Standard are encouraged to investigate the possibility of applying the most recent editions of the standards listed in Annex A.

3 REQUIREMENTS

3.1 Constituents

3.1.1 In the preparation of Indian curry powder any of the spices and condiments listed in IS 1877 may be used. These shall be listed in the descending order of their proportion on the container. Edible starch and edible common salt may be added while preparing the material and declarations to these effect shall be made. The quantities of spices and condiments shall not be less than 85 percent of the total mass.

3.1.2 Edible common salt (*see* IS 253) shall not be more than 5 percent by weight on dry basis.

3.2 Description

Indian curry powder shall be prepared by grinding clean, dried and sound spices and condiments. It shall be free from dirt, insects, and/or rodent matter and moulds when examined with naked eye (corrected, if necessary for abnormal vision) or with such magnification as may be necessary in any particular case. If the magnification exceeds 10x, this fact shall be stated in the test report. The powder shall also be free from added colouring matter and preservatives other than edible common salt.

3.3 Taste and Flavour

Indian curry powder shall have a fresh and characteristic taste and flavour. It shall have no foreign flavour, rancidity and mustiness.

3.4 The Indian curry powder shall also comply with the requirements given in Table 1 and Table 2.

4 SAMPLING

Representative samples of the material shall be drawn according to the procedure described in IS 13145.

5 TESTS

Tests shall be carried out in accordance with the methods referred to in col (4) of Table 1 and col (7) Table 2.

6 QUALITY OF REAGENTS

Unless specified otherwise, pure chemicals and distilled water (*see* IS 1070) shall be employed in tests.

7 PACKING AND MARKING

7.1 Packing

Indian curry powder shall be packed in clean, sound and dry containers/material made of metal, glass, food grade polymers, paper board. The packing material shall be free from any fungal or insect infestation and should not impart any foreign smell. Each container/package shall be securely closed and sealed.

7.2 Marking

The following particulars shall be legibly and indelibly marked or labelled on each container/package of curry powder:

a) Name and address of the manufacturer or packer;

b) Name of the material;
c) Trade-name or brand name, if any;
d) Batch or code number;
e) Net quantity;
f) Best before date;
g) List of ingredients (constituents) based on its composition by weight, in descending order;
h) Directions for use, if any; and
j) Any other markings required under the *Legal Metrology (Packaged Commodities) Rules,* 2011 and the *Food Safety and Standards (Packaging and Labelling) Regulations,* 2011.

8 BIS CERTIFICATION MARKING

The product(s) conforming to the requirements of this standards may be certified as per the conformity assessment schemes under the provisions of the *Bureau of Indian Standards Act*, 2016 and the Rules and Regulations framed thereunder, and the products may be marked with the Standard Mark.

Table 1 Requirements for Indian Curry Powder
(*Clause* 3.4)

Sl No. (1)	Characteristic (2)	Requirement (3)	Method of Test, Ref to (4)
i)	Moisture, percent by weight, *Max*	10.0	**9** of IS 1797
ii)	Volatile oil, percent (*v/w*) on dry basis, *Min*	0.25	**15** of IS 1797
iii)	Non-volatile ether extract, percent by weight on dry basis, *Min*	10.0	**14** of IS 1797
iv)	Total ash (on dry basis), percent by weight on dry basis, *Max*	12.0	**6** of IS 1797
v)	Acid insoluble ash, percent by weight on dry basis, *Max*	1.0	**8** of IS 1797
vi)	Crude fibre, percent by weight on dry basis, *Max*	15.0	**13** of IS 1797
vii)	Lead (as Pb), mg/kg (on dry basis), *Max*	10.0	A of IS 13545
viii)	Cadmium, mg/kg (on dry basis), *Max*	Not detectable	Annex B
ix)	Arsenic, mg/kg (on dry basis), *Max*	Not detectable	Annex C
x)	Total aflatoxin, µg/kg, *Max*	30	IS 16287
xi)	Aflatoxin B1, µg/kg, *Max*	15	IS 16287

Table 2 Microbiological Requirements

(*Clause* 3.4)

Sl No.	**Characteristic**	**Requirement**				**Method of Test, Ref to IS or ISO**
		Sampling Plan [1]		**Limit (cfu/g)**		
		n	***c***	***m***	***M***	
(1)	(2)	(3)	(4)	(5)	(6)	(7)
i)	*Salmonella*	5	0	Absent/25 g	NA	IS 5887 (Part 3)/ Sec 1)
ii)	*Bacillus cereus*	5	0	1 x 10^3	1 x 10^4	IS 5887 (Part 6)
iii)	Sulphite Reducing Clostridia (SRC)	5	2	1 x 10^2	1 x 10^3	ISO 15213

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1) For sampling plan see Annex D.

ANNEX A

(*Clause* 2)

LIST OF REFERRED STANDARDS

IS No./Other Publications	*Title*
IS 253 : 2014	Specification for common salt (*fourth revision*)
IS 265 : 2021	Hydrochloric acid — Specification (*fifth revision*)
IS 1070 : 1992	Reagent grade water (*third revision*)
IS 1797 : 2017	Spices and condiments — Method of test (*third revision*)
IS 1877 : 1985	Spices and condiments — Botanical nomenclature (*third revision*)
IS 13145 : 2014	Spices and condiments — Methods of sampling (*second revision*)
IS 13545 : 2018	Garam Masala — Specification (*first revision*)
IS 5887 (Part 3/ Sec 1) : 2020/ISO 6579-1 : 2017	Methods for detection of bacteria responsible for food poisoning: Part 3 General guidance on methods for the detection of *Salmonella spp.* (*third revision*)
IS 5887 (Part 6) : 2012/ISO 7932 : 2004	Microbiology of food and animal feeding stuffs — Horizontal method for the enumeration of presumptive *Bacillus cereus*: Part 6 colony — Count technique at 30 °C (*first revision*)
ISO 15213 : 2003	Microbiology of food and animal feeding stuffs — Horizontal method for the enumeration of sulfite-reducing bacteria growing under anaerobic conditions
IS 16287 : 2015	Foodstuffs — Determination of aflatoxin B1, and the total content of aflatoxins B1, B2, G1 and G2 in cereals, nuts and derived products — High performance liquid chromatographic method

ANNEX B

[*Table* 1, *Sl No.* (viii)]

DETERMINATION OF CADMIUM CONTENT

B-1 PRINCIPLE

The samples are dissolved in acid or digested in a mixture of sulphuric and nitric acid. The cadmium in solution is determined by conventional atomic absorption spectroscopy.

B-2 GENERAL PRECAUTIONS

Because of the minute amounts of metals involved, special care shall be taken to reduce the reagent blanks to as low a value as possible and to avoid contamination during the test. All apparatus should be thoroughly cleaned with a mixture of hot dilute acids (1 part hydrochloric acid, 1 part concentrated nitric acid, and 3 parts water) followed by thorough washing with water immediately before use.

B-3 APPARATUS

B-3.1 Kjeldahl Flasks — of silica or borosilicate glass fitted with an extension to the neck by means of a B24 ground joint, as shown in Fig. 1. The extension serves to condense the fumes and carries a tap funnel through which the reagents are introduced.

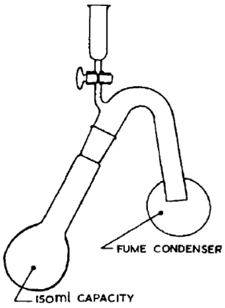

FIG. 1 MODIFIED KJELDAHL FLASK (OPEN TYPE)

B-3.2 Atomic Absorption Spectrophotometer — Any instrument operating in the absorption mode may be used, providing it has facilities for the selection of the required oxidant/fuel combination from a choice of air, argon, nitrous oxide, hydrogen and acetylene and has a wavelength range from 180 nm to 600 nm.

A hydride generation vessel accessory is also required and is available from all the major commercial manufacturers of atomic absorption equipment. For operations in emission mode and measurements of absorption involving the generation of a gaseous hydride, a potentiometric recorder is necessary, preferably a multi-range type covering the range 1 mV to 20 mV.

B-4 REAGENTS

B-4.1 Reagents shall be of an order of purity higher than accepted analytical reagent grade quality. Metal-free water (*see* below) shall be used throughout:

a) *Nitric acid* — sp gr 1.42
b) *Sulphuric acid* — 98 percent
c) *Hydrochloric acid* — 5 N solution prepared by dilution of reagent (d) with metal-free distilled water
d) *Water* — metal-free. Distilled water may be re-distilled from an all-glass apparatus or may he passed down a column or cation exchange resin. For example, Amberlite IR 120 (H)
e) *Sodium borohydride pellets*

B-4.2 Standards

Use commercially available standard solutions or prepare solutions as follows:

a) *Standard cadmium solution* — Dissolve 2.282 g $3CdS0_4.8H_2O$ in distilled water, dilute to 1 000 ml at 20 °C with water in a one-mark graduated flask. Dilute 10.0 ml of this solution to 500 ml at 20 °C with water in a one-mark graduated flask. One milli litre contains 20 µg Cd.

B-5 PREPARATION OF TEST SOLUTIONS

Accurately weigh about 2.5 g of the sample and dissolve in a mixture of 4 ml of sulphuric acid and 5 ml of hydrochloric acid. Transfer the solution to a 50 ml one-mark graduated flask. Dilute to the mark with water. Call this 'Solution A'.

Prepare a reagent blank using the same quantities of reagents as used in the sample digestion.

B-6 MEASUREMENT OF CADMIUM BY ATOMIC ABSORPTION

B-6.1 Preparation of Calibration Curve Solutions

To a series of 100 ml one-mark volumetric flasks, pipette 0 ml, 1 ml, 2 ml, 3 ml, 4 ml and 5 ml of the appropriate standard solution and dilute 10 ml about 50 ml. Add 8 ml concentrated sulphuric acid and 10 ml concentrated hydrochloric acid. Shake to dissolve. When solution is complete, dilute the mark with metal free water. These solutions then contain 0 µg per ml, 0.2 µg per ml, 0.4 µg per ml, 0.6 µg per ml, 0.8 µg per ml and 1.0 µg per ml of cadmium.

B-6.2 Instrumental Conditions

Select the wavelength and gas to be used for the particular element under consideration from the table below:

Element	*Wavelength (nm)*	*Gases*
Cadmium	228.8	Air/acetylene

The recommended settings for the various instrumental parameters vary from model to model, and certain parameters require optimization at the time of use to obtain the best results. Instruments should therefore be adjusted as described in the manufacturer's instructions using the type of flame and wavelength settings specified above.

B-6.3 Procedure

Set the atomic absorption spectrophotometer to the appropriate conditions. Aspirate the strongest standard containing the element to be determined and optimize the instrument settings to give full-scale or maximum deflection on the chart recorder. Measure the absorbances of the other standards and plot a graph showing the net absorbance against the concentration of the element in the standard solution. Aspirate the Solution A obtained from dissolution or the wet oxidation of the sample and the corresponding blank solution and determine the net absorbance. Using the graph prepared above, determine the concentration of the element in the sample solution;

Element in the sample, mg/kg =

$$\frac{Concentration\ of\ element\ (\mu g/ml)\ \times 50}{Mass\ of\ sample\ taken (g)}$$

ANNEX C

[*Table* 1, *Sl No.* (ix)]

DETERMINATION OF ARSENIC CONTENT

C-1 PRINCIPLE

The samples are dissolved in acid or digested in a mixture of sulphuric and nitric acid. The cadmium in solution is determined by conventional atomic absorption spectroscopy.

C-2 GENERAL PRECAUTIONS

Because of the minute amounts of metals involved, special care shall be taken to reduce the reagent blanks to as low a value as possible and to avoid contamination during the test. All apparatus should be thoroughly cleaned with a mixture of hot dilute acids (1 part hydrochloric acid, 1 part concentrated nitric acid, and 3 parts water) followed by thorough washing with water immediately before use.

C-3 APPARATUS

C-3.1 Kjeldahl Flasks — of silica or borosilicate glass fitted with an extension to the neck by means of a B24 ground joint, as shown in Fig. 1. The extension serves to condense the fumes and carries a tap funnel through which the reagents are introduced.

C-3.2 Atomic Absorption Spectrophotometer — Any instrument operating in the absorption mode may be used, providing it has facilities for the selection of the required oxidant/fuel combination from a choice of air, argon, nitrous oxide, hydrogen and acetylene and has a wavelength range from 180 nm to 600 nm.

A hydride generation vessel accessory is also required and is available from all the major commercial manufacturers of atomic absorption equipment. For operations in emission mode and measurements of absorption involving the generation of a gaseous hydride, a potentiometric recorder is necessary, preferably a multi-range type covering the range 1 mV to 20 mV.

C-4 REAGENTS

C-4.1 Reagents shall be of an order of purity higher than accepted analytical reagent grade quality. Metal-free water (*see* below) shall be used throughout:

a) *Nitric acid* — sp gr 1.42
b) *Sulphuric acid* — 98 percent
c) *Hydrochloric acid* — 5 N solution prepared by dilution of reagent (d) with metal-free distilled water
d) *Water* — metal-free. Distilled water may be re-distilled from an all-glass apparatus or may he passed down a column or cation exchange resin. For example, Amberlite IR 120 (H)
e) *Sodium borohydride pellets*

C-4.2 Standards

Use commercially available standard solutions or prepare solutions as follows:

a) *Standard arsenic solution* — Dissolve 1.320 g of arsenous oxide. As_2O_3 by warming at a temperature not exceeding 60 °C with 14 ml of 5 N sodium hydroxide solution in a 100 ml beaker. Cool, add 0.2 ml of phenolphthalein indicator and neutralize with 6 N sulphuric acid. Transfer the solution to a 1 000 ml one-mark graduated flask containing 10 g of sodium hydrogen carbonate dissolved in water, washing out the beaker with water. Dilute to the mark with water at 20 °C and mix. Dilute 5 ml of this solution to 1 000 ml at 20 °C with water in a one-mark graduated flask as required. 1 ml contains 5 µg As.

C-5 MEASUREMENT OF ARSENIC AND ANTIMONY BY ATOMIC ABSORPTION HYDRIDE TECHNIQUE

Arsenic is determined after preparation of its volatile hydrides which are collected either in the generation vessel itself or, in some designs, in a rubber balloon attached to the vessel. The gases are then expelled with argon into a hydrogen flame.

C-5.1 Preparation of Calibration Curve Solution

Into a series of 100-ml one-mark volumetric flasks add from a burette, 0 ml, 1 ml, 2 ml, 3 ml, 4 ml and 5 ml of standard arsenic solution and dilute to about 50 ml with distilled water. Add 8 ml concentrated sulphuric acid. Shake to dissolve, and when solution is complete, dilute to the mark with distilled water.

C-5.2 Instrumental Conditions

Using the atomic absorption spectrophotometer with the appropriate hollow cathode or electrodeless discharge lamp, select the wavelength for either arsenic (1 937 nm).

C-5.3 Procedure

Measure 5.0 ml of the strongest standard into the generation vessel, add 25 ml water and 2 ml 5 N hydrochloric acid. Stopper the vessel and expel any air as described in the maker’s instructions, filling the apparatus with argon. Isolate the vessel from the atomizer using the by-pass valve. Remove the atomizer and then quickly add 1 pellet of sodium borohydride weighing approximately 0.2 g and replace the stopper. Ensure that all the joints are secure.

When the reaction slows (20 sec to 30 sec) open the appropriate taps to allow argon to drive the generated hydride into the flame. When the hydride has all been expelled as shown by the recorder trace, return the taps to their original position and empty the vessel.

Optimize the instrument setting to give full scale deflection for the strongest standard. Measure the other standards, the sample and the blank solution using the same procedure.

Plot the graph relating peak height on the recorder to concentration of the arsenic in the standards. Using the net absorbance of the sample, read the graph the concentration of arsenic in the solution.

C-5.6 Calculation

Arsenic in the sample, mg/kg =

$$\frac{Concentration\ of\ arsenic\ (\frac{\mu g}{ml})}{Mass\ of\ sample\ taken\ (g)} \times 50$$

ANNEX D

(*Table* 2)

SAMPLING PLAN FOR MICROBIOLOGICAL REQUIREMENTS

D-1 Sampling Plan for Microbiological Requirements

The terms n, c, m and M used in this standard have the following meaning:

n = Number of units comprising a sample;

c = Maximum allowable number of units having microbiological counts above *m* for 2- class sampling plan and between *m* and *M* for 3 - class sampling plan;

m = Microbiological limit that separates unsatisfactory from satisfactory in a 2- class sampling plan or acceptable from satisfactory in a 3-class sampling plan; and

M = Microbiological limit that separates unsatisfactory from satisfactory in a 3 - class sampling plan.

D-2 Interpretation of Results:

2-Class Sampling Plan (*where n, c and m are specified*)	*3-Class Sampling Plan* (*where n, c, m and M are specified*)
1) Satisfactory, if all the values observed are $\leq m$	1) Satisfactory, if all the values observed are $\leq m$
2) Unsatisfactory, if one or more of the values observed are > *m*	2) Acceptable, if a maximum of *c* values are between *m* and *M*
	3) Unsatisfactory, if one or more of the values observed are > *M* or more than prescribed *c* values are > *m*

ANNEX E

(*Foreword*)

COMMITTEE COMPOSITION

Spices and Condiments Sectional Committee, FAD 09

Organization	*Representative*(*s*)
Spices Board India, Kochi	DR A.B. REMA SHREE **(*Chairperson*)**
AB Mauri India Pvt Ltd, Kochi	SHRI PRAKASH NAMBOODIRI
All India Spices Exporters Forum, Cochin	SHRI CHERIAN XAVIER MS PRIYAMVADA NILAYANGOD (*Alternate*)
CSIR - Central Food Technological Research Institute, Mysore	DR M. MADHAVA NAIDU SHRI NAGARAJAN S. (*Alternate*)
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Consumer Coordination Council, Noida	SHRI RAMJI BHAI MAVANI
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Directorate of Arecanut and Spices Development, Calicut	DR HOMEY CHERIYAN DR FEMINA (*Alternate*)
Directorate of Marketing and Inspection, Faridabad	SHRI SHIVNANDAN SHRI RAHUL SINGH (*Alternate*)
Export Inspection Council of India, New Delhi	SHRI WASI ASGHAR SHRI SHASHI PRAKASH TRIPATHI (*Alternate*)
Food Safety and Standards Authority of India, New Delhi	ADVISOR
ICAR - Indian Institute of Spices Research, Kozhikode	DR N. K. LEELA DR E. JAYASHREE (*Alternate*)
ICMR - National Institute of Nutrition, Hyderabad	DR S. SREENIVASA REDDY DR PARAS SHARMA (*Alternate*)
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National Research Centre on Seed Spices, Ajmer	DR S.N. SAXENA DR B.K. MISHRA (*Alternate*)

Organization	*Representative*(*s*)
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Spices Board India, Kochi	SHRI RAMESH BABU N MS C.M. SRI LATHA (*Alternate*)
Synthite Industries Pvt Ltd, Kolenchery	SHRI SHUBU K. CHITHAJAN SHRI ANIL KUMAR (*Alternate*)
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Amendments are issued to standards as the need arises on the basis of comments. Standards are also reviewed periodically; a standard along with amendments is reaffirmed when such review indicates that no changes are needed; if the review indicates that changes are needed, it is taken up for revision. Users of Indian Standards should ascertain that they are in possession of the latest amendments or edition by referring to the website-www.bis.gov.in or www.standardsbis.in.

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Amendments Issued Since Publication

Amend No.	Date of Issue	Text Affected

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